

Nor Hazliana Harun<sup>1</sup>, Rabiatal Basria S.M.N. Mydin<sup>1</sup>, Srimala Sreekantan<sup>2</sup>, Khairul Arifah Saharudin<sup>2</sup>, Khor Yong Ling<sup>2</sup>, Norfatehah Basiron<sup>2</sup>, Azman Seeni<sup>1,3</sup>

<sup>1</sup>Oncological and Radiological Sciences Cluster, Advanced Medical & Dental Institute, Universiti Sains Malaysia, Penang, Malaysia

<sup>2</sup>School of Materials & Mineral Resources Engineering, Engineering Campus, Universiti Sains Malaysia, Penang, Malaysia

<sup>3</sup>Malaysian Institute of Pharmaceuticals and Nutraceuticals (IPHARM), National Institute of Biotechnology Malaysia, Penang, Malaysia

\*Corresponding author:  
Rabiatal Basria S.M.N. Mydin  
E-mail: [rabiatalbasria@usm.my](mailto:rabiatalbasria@usm.my)

Received 01 Apr 2018.  
Revised 01 June 2018.  
Accepted 25 July 2018.  
Published Online 15 Aug 2018

## Antibacterial activity of heterogeneous TiO<sub>2</sub> and ZnO nanoparticles against Gram-positive and Gram-negative bacterial pathogens

**Abstract**—Hospital-acquired infections (HAIs) are responsible for over 40% of cases in acute-care hospitals and commonly associated with catheters-associated urinary tract infections (CAUTIs). Current nanotechnology approach focus on improving the aseptic procedures for medical devices and manage the HAIs risk. TiO<sub>2</sub> and ZnO nanoparticles (NPs) have been widely reported independently, to have a photocatalytic killing potential. The present study evaluates the antibacterial activity of heterojunction between TiO<sub>2</sub> and ZnO NPs on several types bacterial pathogens model including *Staphylococcus aureus*, *Enterococcus faecalis*, *Escherichia coli*, *Klebsiella pneumoniae*, and *Pseudomonas aeruginosa*. The antibacterial screening test on TiO<sub>2</sub>/ZnO nanoparticles (NPs) were done under dark and light conditions with different molar ratio 25T75Z, 50T50Z and 75T25Z according to Clinical Laboratory Standards Institute (CLSI) guidelines MO2-A11. ZnO and TiO<sub>2</sub>/ZnO (25T75Z and 50T50Z) NPs at the highest concentration (1000µg/µL) showed mean diameters of the zones of inhibition (mm); (12.5 ± 0.58), (12.13 ± 0.85), and (7.25 ± 1.44) in dark condition. Increment in inhibition zones was obtained under light condition; (21.38 ± 0.48), (17.50 ± 1.0), and (12.38 ± 1.80). Findings from this study highlights the heterogeneous TiO<sub>2</sub> and ZnO NPs could become a promising bacteriostatic and/or bactericidal agent to combat against the HAIs.

**Keywords** — TiO<sub>2</sub>/ZnO nanoparticles, Hospital-acquired infections, Antibacterial activity, Bacteriostatic agent, Bactericidal agent, Biomedical Product, Biomaterial, Nanomaterial

### 1 INTRODUCTION

Healthcare-associated infections (HAIs) caused by Gram-positive and Gram-negative bacterial pathogens are significant economic and public health burdens globally, having an estimated level of 6.4% and 1,000,000 reported cases annually [1]. The most common HAI pathogens include Gram-positive bacteria, such as *Staphylococcus aureus*, *Staphylococcus epidermidis* and *Enterococcus faecalis*, and Gram-negative bacteria, including *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis* and *Pseudomonas aeruginosa*, and yeasts, such as *Candida* sp. [2]. HAI is closely related to medical device contamination which poses high patient morbidity and mortality cases [3]. Rutala et al. (2013) studied a rational approach in cleaning, disinfection and sterilisation techniques for patient care items and medical devices developed by Earle H. Spaulding for the last four decades as one of the strategies to reduce HAIs [4]. Failure to provide a proper disinfection or sterilisation on

medical devices may lead to the introduction of pathogenic infection, thereby risking patients' lives.

Advancement in nanotechnology can provide new approaches in controlling HAI risks. The physicochemical properties of metal oxides NPs, such as TiO<sub>2</sub> and ZnO, have substantial important uses in several pharmaceutical industries. A number of studies have indicated the widely uses of TiO<sub>2</sub> and ZnO in therapeutic applications such as biosensing [5], imaging labels [6,7] and drug delivery [8,9]. These NPs also present unique properties such as cytocompatibility, biocompatibility, chemical stability, environmental-friendliness, wide band gap and relatively low cost [10, 11]. Moreover, TiO<sub>2</sub> and Zn NPs have been widely studied as antibacterial agents owing to their photocatalytic activities [12-15]. In fact, several efforts have been successful in improving the photocatalytic capabilities and synergistic effects of metal oxide NPs, such as CuO/ZnO, cerium oxide/yttrium

oxide and Ag orthophosphate ( $\text{Ag}_3\text{PO}_4/\text{TiO}_2/\text{Fe}_3\text{O}_4$ ) NPs [16,17]. Thus, the present study investigates the antibacterial activity of  $\text{TiO}_2/\text{ZnO}$  NPs with different molar ratios against the selected Gram-positive and -negative bacteria under both dark and visible light conditions.

## 2 METHODOLOGY

### 2.1 $\text{TiO}_2$ / ZnO NPs synthesis and characterization

Titanium (IV) isopropoxide 97% and Zinc acetate dehydrate 98% were supplied by Sigma Aldrich. The  $\text{TiO}_2/\text{ZnO}$  NPs photocatalyst of different molar ratio of  $\text{TiO}_2$  and ZnO was synthesis by sol-gel process with molar ratio at 25T75Z, 50T50Z and 75T25Z. Commercial  $\text{TiO}_2$  P25, 100T and 100Z represent as control counterpart. The ZnO NPs were prepared as follows. ZnO sol was prepared by adding 0.114 mole Zinc acetate dehydrate (ZAD) into ethanol. The solution was stirred for 5 min in heated (70 °C) water bath to obtain the precursor solution. Next, continuous stirring for 5 h were carried out until transparent solution is obtained. Then, deionized water was added drop wise and stirred for 10 min. On the other hand, the  $\text{TiO}_2$  NPs were prepared by dissolving Titanium Isopropoxide (TTIP) in ethanol with the volume ratio of 1:4 and stirred for 30 min. Then, deionised water was added dropwise and stirred for 3 h. After  $\text{TiO}_2$  and ZnO sols are synthesized separately, the sols are mixed to different ratios and stirred continuously for 1h. The milky white suspension was centrifuged, washed with ethanol until white sediment was observed. The resultant sediment was oven dried overnight at 80 °C. The dried white precipitate was pulverized with pestle and mortar and calcined in air at 500 °C for 2h. The morphology of the  $\text{TiO}_2/\text{ZnO}$  NPs photocatalyst was further characterized using Field Emission Scanning Electron Microscope (FESEM) model of LEO GEMINI.

### 2.2 Photocatalytic activity

The photocatalytic activity  $\text{TiO}_2/\text{ZnO}$  NPs photocatalyst were studied by degradation of methylene blue (MB) solution under sunlight. 10 mg of each catalyst was suspended into 40 mL MB aqueous solution (3 ppm). The mixture was poured into a glass petri dish and kept in a dark environment for 30 min to allow it to attain an equilibrium adsorption state. It was then put under sunlight for a set of irradiation time. After the degradation, the mixture was withdrawn and

the catalyst was separated from the suspensions by a centrifuge. The determination of MB concentration was performed using a Cary50 UV-vis spectrophotometer (Varian Corp) by measuring the absorption of MB at 644 nm.

### 2.3 Antibacterial Disc Susceptibility Tests

*S. aureus* ATCC 25923, *E. faecalis* ATCC 29212, *E. coli* ATCC 25922, *K. pneumoniae* ATCC 700603 and *P. aeruginosa* ATCC 27853 were tested. They were cultured aerobically at 37 °C on Luria–Bertani agar plates (Merck, Germany) for 24 h. Positive control was represented by the standard antibiotic and negative control discs (sample diluent) from 10% dimethyl sulfoxide (DMSO) (Sigma-Aldrich). Test samples were prepared in 10% DMSO as diluent with different concentrations (100, 200, 500, 800 and 1000 µg/µl). The disc diffusion assay of  $\text{TiO}_2/\text{ZnO}$  NPs was conducted according to the guidelines of CLSI MO2-A11 (Clinical and Laboratory Standard Institute MO2-A11) under dark and light conditions with visible light intensity of 5.70  $\text{w/m}^2$  [18]. All tests were done in triplicate.

## 3 RESULTS AND DISCUSSION

FESEM images of NPs are shown in Figure 1a–d. Most of the NPs photocatalyst were spherical in shape. Nevertheless, in certain regions of 25T75Z (Figure 1c) and 100Z (Figure 1d), rod structures were visible (marked in red square). The average particle size of the spherical shape was 91.35 nm, while the nanorod diameter and length were 66.13 and 409.03 nm, respectively.

The photocatalytic activity of 100T, 25T75Z, 50T50Z, 75T25Z and 100Z were evaluated by the photocatalytic decolourisation of methylene blue (MB) under sunlight irradiation (Figure 2). MB concentration was decreases with the increase in irradiation time. After 180 min of sunlight exposure, 100T, 25T75Z and 100Z displayed ~98% degradation, whilst 75T25Z and 50T50Z showed only 88% and 50% degradation, respectively. The results suggested that the improvement in photocatalytic activity in bare ZnO was contributed by less efficient radioactive recombination at UV emission. Meanwhile, for  $\text{TiO}_2/\text{ZnO}$  NPs photocatalyst, especially 25T75Z, enhancement was related to intrinsic defects and charge separation due to improved interface between ZnO and  $\text{TiO}_2$ .

Additionally, the photocatalytic degradation of MB for the entire photocatalyst was quantitatively investigated through a pseudo-first-order kinetic model, and the observed first-order

rate constant (k) was determined. The values of the calculated k constants are presented in Figure 3. Kinetic data revealed that the degradation rate constants for 100T, 100Z, 25T75Z, 50Z50T and 75T25Z under sunlight irradiation were  $1.55 \times 10^{-2}$ ,  $1.71 \times 10^{-2}$ ,  $1.80 \times 10^{-2}$ ,  $0.4 \times 10^{-2}$  and  $1.13 \times 10^{-2} \text{ min}^{-1}$ , respectively. Thus, the degradation rate showed in an increasing order, that is,  $50T50Z < 75T25Z < 100T < 100Z < 25T75Z$ . This result indicated that the presence of  $\text{TiO}_2$  and ZnO (25T75Z) and the coexistence of zincite phase resulted in highest activity for NPs.

Meanwhile, the antibacterial profiles of  $\text{TiO}_2/\text{ZnO}$  NPs in terms of the differential molar ratios, concentrations and conditions against *S. aureus* are shown in Figure 4. The effect of  $\text{TiO}_2/\text{ZnO}$  NPs on *S. aureus* showed large inhibition zones at 1000  $\mu\text{g}/\mu\text{l}$  with the presence of visible light, that is,  $21.38 \pm 0.48$ ,  $17.50 \pm 1.0$  and  $12.38 \pm 1.80$ , for 100Z, 25T75Z and 50T50Z NPs, respectively. *S. aureus* also showed inhibition zones even under dark condition with the maximum inhibition zones of  $12.5 \pm 0.58$ ,  $12.13 \pm 0.85$  and  $7.25 \pm 1.44$  for 100Z, 25T75Z and 50T50Z NPs, respectively. Jesline et al. (2014) reported similar finding on the antibacterial activities of  $\text{TiO}_2$  and ZnO NPs against *S. aureus* [19]. The ability of ZnO NPs and the heterogeneity of ZnO-based compounds are well-known excellent antibacterial agents. Thus, high ZnO content in the heterogeneous  $\text{TiO}_2/\text{ZnO}$  samples resulted in large inhibition zones in both conditions. The result indicated the bacteriostatic capability of 25T75Z and 50T50Z NPs against *S. aureus* in both conditions.

The bacteriostatic activity of ZnO and  $\text{TiO}_2$  NPs photocatalyst under both conditions may be attributed to photocatalytic activity, which produced superoxide anion radicals. These superoxide anion radicals reacted with  $\text{H}^+$  ions to form  $\text{HO}_2^-$  radicals, which in turn reacted with electrons and  $\text{H}^+$ -yielding  $\text{H}_2\text{O}_2$  molecules. Subsequently,  $\text{H}_2\text{O}_2$  molecules penetrated the cell membrane and caused the death of *S. aureus*. The generation of O species, such as  $\text{H}_2\text{O}_2$ ,  $\text{O}_2^-$  and  $\text{OH}^*$ , disrupted the bacterial membrane structure and killed the bacteria [20]. Several studies also reported ROS generation activity under dark conditions [21,22]. Therefore, our study confirmed that ROS generation under dark and visible light conditions resulted in phototoxic effects that inhibited or killed *S. aureus*. In the *S. aureus* bacterial cell wall, it consisting several layers of peptidoglycan. The porous structure of

the interconnected peptidoglycan layer and interaction between positively charged NPs with negatively charged teichoic acid of *S. aureus* allowing diffusion of NPs into the bacterial membrane [26]. However, no inhibition zones were observed in the Gram-negative bacteria tested in this study under both (light and dark) conditions. This phenomenon explain due to the greater complexity of the double membranes of the Gram-negative bacteria which composed of asymmetric bilayers of phospholipid and lipopolysaccharides. It is apparent that the outer membrane serve as selective barriers that provide an extra layer of protection for diffusion or penetration of NPs [23-25].

#### 4 CONCLUSION

The bactericidal activity of heterogeneous  $\text{TiO}_2/\text{ZnO}$  NPs is not only dependent on the concentration and type of bacteria but also on the photocatalytic activity via the generation of reactive oxygen species, such as superoxide anion radical, hydroxyl radical and  $\text{H}_2\text{O}_2$ . The potential bacteriostatic agents of  $\text{TiO}_2/\text{ZnO}$  NPs with molar ratios of 25T75Z and 50T50Z against HAIs requires further study.

#### ACKNOWLEDGEMENT

The authors are thankful to the Ministry of Education (MOE) Malaysia for funding this work under Transdisciplinary Research Grant Scheme (TRGS) grant no. 6769003 and Universiti Sains Malaysia (USM) for providing the necessary facilities to carry out the research work.

#### CONFLICTS OF INTEREST

The authors report no conflicts of interest. The authors are responsible for the content and writing of the paper.

#### REFERENCES

- [1] HPA (2012a). English National Point Prevalence Survey on Healthcare- Associated Infections and Antimicrobial Use, 2011: Preliminary Data. London: Health Protection Agency.
- [2] Donlan RM. Biofilms and device-associated infections. *Emerging infectious diseases*. 2001 Mar;7(2):277.
- [3] Donlan RM. Biofilms on central venous catheters: is eradication possible?. In *Bacterial biofilms 2008* (pp. 133-161). Springer Berlin Heidelberg.
- [4] Rutala WA, Weber DJ. Disinfection and sterilization: an overview. *American journal of infection control*. 2013 May 31;41(5):S2-5.
- [5] Rajh T, Dimitrijevic NM, Bissonnette M, Koritarov T, Konda V. Titanium dioxide in the service of the

- biomedical revolution. *Chemical reviews*. 2014 Aug 29;114(19):10177-216.
- [6] Hong H, Shi J, Yang Y, Zhang Y, Engle JW, Nickles RJ, Wang X, Cai W. Cancer-targeted optical imaging with fluorescent zinc oxide nanowires. *Nano letters*. 2011 Aug 10;11(9):3744-50.
- [7] Liu Y. Application of Gadolinium-Doped Zinc Oxide Quantum Dots for Magnetic Resonance and Fluorescence Imaging. In *Multifunctional Nanoprobes 2018* (pp. 65-79). Springer, Singapore.
- [8] Ansari SA, Husain Q, Qayyum S, Azam A. Designing and surface modification of zinc oxide nanoparticles for biomedical applications. *Food and Chemical Toxicology*. 2011 Sep 1;49(9):2107-15.
- [9] Yuan Q, Hein S, Misra RD. New generation of chitosan-encapsulated ZnO quantum dots loaded with drug: synthesis, characterization and in vitro drug delivery response. *Acta Biomaterialia*. 2010 Jul 1;6(7):2732-9.
- [10] Weir A, Westerhoff P, Fabricius L, Hristovski K, Von Goetz N. Titanium dioxide nanoparticles in food and personal care products. *Environmental science & technology*. 2012 Feb 8; 46(4):2242-50.
- [11] Fan Z, Lu JG. Zinc oxide nanostructures: synthesis and properties. *Journal of nanoscience and nanotechnology*. 2005 Oct 1;5 (10):1561-73.
- [12] Mao, Y., Park, T. J., & Wong, S. S. (2005). Synthesis of classes of ternary metal oxide nanostructures. *Chemical Communications*, (46), 5721-5735.
- [13] Yalcinkaya F, Lubasova D. Quantitative evaluation of antibacterial activities of nanoparticles (ZnO, TiO<sub>2</sub>, ZnO/TiO<sub>2</sub>, SnO<sub>2</sub>, CuO, ZrO<sub>2</sub>, and AgNO<sub>3</sub>) incorporated into polyvinyl butyral nanofibers. *Polymers for Advanced Technologies*. 2017 Jan 1;28(1):137-40.
- [14] Jašková V, Hochmannová L, Vytřasová J. TiO<sub>2</sub> and ZnO nanoparticles in photocatalytic and hygienic coatings. *International journal of photoenergy*. 2013 Feb 24;2013.
- [15] Talebian N, Doudi M, Mogoei H. Antibacterial activities of sol-gel derived ZnO-multilayered thin films: p-NiO heterojunction layer effect. *Journal of Sol-Gel Science and Technology*. 2015 Jun 1;74(3):650-60.
- [16] Widiarti N, Sae JK, Wahyuni S. Synthesis CuO-ZnO nanocomposite and its application as an antibacterial agent. In *IOP Conference Series: Materials Science and Engineering 2017 Feb* (Vol. 172, No. 1, p. 012036). IOP Publishing.
- [17] Xu JW, Gao ZD, Han K, Liu Y, Song YY. Synthesis of magnetically separable Ag<sub>3</sub>PO<sub>4</sub>/TiO<sub>2</sub>/Fe<sub>3</sub>O<sub>4</sub> heterostructure with enhanced photocatalytic performance under visible light for photoinactivation of bacteria. *ACS applied materials & interfaces*. 2014 Aug 27;6(17):15122-31.
- [18] Edition AS. CLSI document M02-A11. Wayne, PA: Clinical and Laboratory Standards Institute. 2012;32(1):76.
- [19] Jesline A, John NP, Narayanan PM, Vani C, Murugan S. Antimicrobial activity of zinc and titanium dioxide nanoparticles against biofilm-producing methicillin-resistant *Staphylococcus aureus*. *Applied Nanoscience*. 2015 Feb 1;5(2):157-62.
- [20] Joe A, Park SH, Shim KD, Kim DJ, Jhee KH, Lee HW, Heo CH, Kim HM, Jang ES. Antibacterial mechanism of ZnO nanoparticles under dark conditions. *Journal of Industrial and Engineering Chemistry*. 2017 Jan 25;45:430-9.
- [21] Hirota K, Sugimoto M, Kato M, Tsukagoshi K, Tanigawa T, Sugimoto H. Preparation of zinc oxide ceramics with a sustainable antibacterial activity under dark conditions. *Ceramics International*. 2010 Mar 31;36(2):497-506.
- [22] Kirkinezos IG, Moraes CT. Reactive oxygen species and mitochondrial diseases. In *Seminars in cell & developmental biology 2001 Dec 31* (Vol. 12, No. 6, pp. 449-457). Academic Press.
- [23] Clifton LA, Skoda MW, Daulton EL, Hughes AV, Le Brun AP, Lakey JH, Holt SA. Asymmetric phospholipid: lipopolysaccharide bilayers; a Gram-negative bacterial outer membrane mimic. *Journal of The Royal Society Interface*. 2013 Dec 6;10(89):20130810.
- [24] Santos RS, Figueiredo C, Azevedo NF, Braeckmans K, De Smedt SC. Nanomaterials and molecular transporters to overcome the bacterial envelope barrier: Towards advanced delivery of antibiotics. *Advanced drug delivery reviews*. 2017 Dec 14. Egan, A. J. (2018). Bacterial outer membrane constriction. *Molecular microbiology*.
- [25] Bajaj H, Acosta Gutierrez S, Bodrenko I, Mallocci G, Scorciapino MA, Winterhalter M, Ceccarelli M. Bacterial outer membrane porins as electrostatic nanosieves: exploring transport rules of small polar molecules. *ACS nano*. 2017 May 12;11(6):5465-73
- [26] Auer GK, Weibel DB. Bacterial cell mechanics. *Biochemistry*. 2017 Jul 11;56(29):3710-24