A'attiyyah AA¹, Wan Afiqah Syahirah WG¹, Nor Azah MA², Kannan TP^{1,3,*}, Suharni M¹, Ahmad A¹

¹School of Dental Sciences Universiti Sains Malaysia, 16150, Kubang Kerian, Kelantan, Malaysia

²Herbal Product Development Laboratory, Natural Products Division, Forest Research Institute of Malaysia, Kepong, 52109, Selangor, Malaysia

³Human Genome Centre, School of Medical Sciences, Universiti Sains Malaysia, 16150, Kubang Kerian, Kelantan, Malaysia

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*Corresponding author: Kannan TP E-mail: kannan@usm.my

Phytochemical Properties and Traditional Uses of Selected Medicinal Plants in Malaysia: A Review

Abstract - Medicinal plants have healing properties and are able to synthesize various chemical compounds. These chemicals (also known as phytochemical compounds) play vital roles in determining the pharmacological properties existing in certain plants. The phytochemical compounds present in plants are associated with primary and secondary constituents. Most of the time, the secondary constituents exhibit the bioactivities in plants such as antimicrobial, antioxidant, antidiabetic, antibacterial and anti-inflammatory properties. Some common medicinal plants that have been used in curing various diseases by traditional practitioners in Malaysia are *Ficus deltoidea* Jack, *Andrographis paniculata*, *Curcuma longa*, *Clinacanthus nutans* and *Eurycoma longifolia* Jack. This review discusses the morphology, phytochemical compounds and phytochemical properties of selected medicinal plants in Malaysia. The plants of focus have been found to possess anti-cancer and anti-diabetic effects. This review, it is hoped will enable Malaysian researchers to explore further on the potential of these plants in investigating new and novel drugs in the future.

Keywords - Malaysia, plants, phytochemical compounds, properties

1 INTRODUCTION

Malaysia is one of the tropical rainforest countries that is rich with its traditional medicines and herbal plants. Herbal plants have been used for many years to cure various diseases including diabetes mellitus, skin rashes, fever, insect and snake bites. As stated by World Health Organization (WHO), each part of the plant is useful and contains active compounds that are important for therapeutic purposes or which are precursors for the synthesis of useful drugs [1]. These active compounds such as tannins, saponins, alkaloids, terpenoids, and flavonoids produce definite steroids physiological actions in human [2]. Some of these active compounds are non-essential towards the plants that produce them [2], even though they act as a powerful medicine for human beings.

Phytochemical compounds can be divided into two categories which are primary and secondary constituents [3]. Primary constituents consist of chlorophylls, proteins, amino acids and common sugars while the secondary constituents consist of terpenoids, alkaloids and phenolic compounds [3]. Terpenoids are chemical compounds that are derived from isoprene molecule (C_5H_8) [4]. Scientifically, the roles of terpenoids in pharmacological activities are anti-inflammatory, anti-bacterial, anti-protozoan, anti-fungal, antiviral, anti-allergen in addition to acting as an immune booster and antineoplastic [3].

Alkaloids were first introduced by Meisner in the beginning of nineteenth century to designate natural compounds that react like a base, alkali, since alkaloids are derived from organic nitrogenous bases. The alkaloids are plant-derived compounds containing nitrogen and can be found in over 20% of plant species [5, 6]. Alkaloids have high impact on human beings in many areas including medical, economics and social affairs [5]. Phenolic compounds or polyphenolic compounds are one of the secondary constituents that can be found in plants and are involved in defence against ultra-violet radiation and aggression by pathogens [7]. Chemically, phenolic compounds contain at least 1 aromatic hydrocarbon ring with 1 or more hydroxyl groups attached [6]. Flavonoids,

coumarins, benzoic acids and tannins can be classified as polyphenolic compounds with the flavonoids as the largest and most diverse group [6]. These secondary constituents are most essential in pharmaceutical developments besides being vital in the adaptations of plants to their environments [8]. Almost all humans in the world prefer traditional plants as a treatment option and around 80 % of the world population rely on traditional medicines for primary health care [9]. In Malaysia, a variety of medicinal plants exist which include Ficus deltoidea, Andrographis paniculata, Clinacanthus nutans and Curcuma longa, Cinnamomum verum. This paper will discuss the phytochemical compounds present in five selected common traditional plants in Malaysia, namely, Ficus deltoidea. Andrographis paniculata, Cinnamomum verum, Clinacanthus nutans and Curcuma longa.

This review was prepared by searching some databases like PubMed, Medline, Scopus, PLoS and Google Scholar using the keywords phytochemical properties, Ficus deltoidea Jack, Andrographis paniculata, Curcuma longa, Cinnamomum verum and Clinacanthus nutans. For each medicinal plant, the general information about phytochemical compounds, their families, the pharmacological properties and their traditional uses are reported. However, the phytochemical properties of these plants are limited only to that reported by Malaysian researchers.

2 COMMON MEDICINAL PLANTS IN MALAYSIA AND THEIR MORPHOLOGY

The plants included in this review are *Ficus deltoidea* (*F. deltoidea*), *Andrographis paniculata* (*A. paniculata*), *Cinnamomum verum* (*C. verum*), *Clinacanthus nutans* (*C. nutans*) and *Curcuma longa* (*C. longa*). Tables I and II give the details of the synonyms, common names and the morphology of these medicinal plants.

Plants	<i>F. deltoidea</i> Jack	A. paniculata	C. nutans	C. verum	C. longa
Family	Moraceae	Acanthaceae	Acanthaceae	Lauraceae	Zingiberaceae
Synonyms	F. deltoidea var. angustifolia (Miq.) Corner, F. deltoidea forma angustissina Corner, F. deltoidea var. arenaria Corner,	<i>Justicia paniculata</i> d	Clinacanthus nutans var. robinsonii Benoist, Clinacanthus burmanni Nees, Justicia nutans Burm. F	Cinnamomum zeylanicum	<i>Curcuma domestica</i> Valeton
	F. deltoidea var. bilobata Corner, F. deltoidea var. borneensis Corner, F. deltoidea forma subhirsuta Corner,				
	F. detoidea var. kunstleri (King) Corner, F. deltoidea var. lutescens (Desf.) Corner, F. deltoidea forma longipedunculata Comer, F. deltoidea forma subsessilis (Miq.) Corner, F. deltoidea var. peltata Corner, F. deltoidea var. recurvate Kochummen and F. detoidea var.				
	trengganuensis Corner.				
ommon	Mas cotek	Hempedu bumi	Belalai daiah	Kulit kavu manis	Kunvit
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able II: Morpl	hology of the selected medicinal plants Morphology	5			
Table II: Morp Plants F. deltoidea	hology of the selected medicinal plants Morphology • Leaf shapes: deltoid, elliptic obov • Leaf lamina: oblong, elliptic, obtri	s /ate, spatulate or rhom! angular, oblanceolate,	poid spatulate, linear and subc	rbicular (12)	
Fable II: Morp Plants deltoidea A. paniculata	hology of the selected medicinal plants Morphology Leaf shapes: deltoid, elliptic obox Leaf lamina: oblong, elliptic, obtri branched annual herbaceous pla grows erect to a height of 30.0-1 Leaf: glabrous, up to 8.0 cm long Stem: dark green with 0.3-1.0 m angles of the younger parts (80)	s vate, spatulate or rhomt angular, oblanceolate, nt 10.0 cm and 2.5 cm broad, pini in height, 2.0-6.0 mm ir	poid spatulate, linear and subc nate, acute apex and entir n diameter, quadrangular	rbicular (12) e margin by having a with longitudinal furrov	smooth edge vs and wings on th
Fable II: Morpi Plants 7. deltoidea A. paniculata C. nutans	hology of the selected medicinal plants Morphology Leaf shapes: deltoid, elliptic obov Leaf lamina: oblong, elliptic, obtri branched annual herbaceous pla grows erect to a height of 30.0-1 Leaf: glabrous, up to 8.0 cm long Stem: dark green with 0.3-1.0 m angles of the younger parts (80) perennial herb/shrub grows around 1.0-3.0 m in height Leaf: pale green, simple, opposit Stem: straight green with white ir	s vate, spatulate or rhomi angular, oblanceolate, nt 10.0 cm and 2.5 cm broad, pini in height, 2.0-6.0 mm ir twith pubescent or sho e, narrowly elliptic oblo itemodes and vertical s	poid spatulate, linear and subc nate, acute apex and entir n diameter, quadrangular n diameter, quadrangular thair branches ng or lanceolate with a dia trips (81)	rbicular (12) e margin by having a with longitudinal furrov meter 0.5-1.5 cm and	smooth edge vs and wings on th 2.5-13.0 cm long
Table II: Morp Plants - deltoidea 1. paniculata 2. nutans 2. verum	hology of the selected medicinal plants Morphology Leaf shapes: deltoid, elliptic obox Leaf lamina: oblong, elliptic, obtri branched annual herbaceous pla grows erect to a height of 30.0-1 Leaf: glabrous, up to 8.0 cm long Stem: dark green with 0.3-1.0 m angles of the younger parts (80) perennial herb/shrub grows around 1.0-3.0 m in height Leaf: pale green, simple, opposit Stem: straight green with white ir small evergreen tropical tree grows up to 10.0-15.0 m tall Leaf: leathery appearance and ut green and shiny adaxially, ovate	s vate, spatulate or rhomi angular, oblanceolate, nt 10.0 cm and 2.5 cm broad, pini in height, 2.0-6.0 mm ir twith pubescent or sho e, narrowly elliptic oblo itemodes and vertical s sually opposite, glabro or ovate-lanceolate, 11	poid spatulate, linear and subc nate, acute apex and entir n diameter, quadrangular nt hair branches ng or lanceolate with a dia trips (81) us on both surfaces while .0 to 16.0 cm long, with pr	rbicular (12) e margin by having a with longitudinal furrov meter 0.5-1.5 cm and binted tips (82)	smooth edge vs and wings on th 2.5-13.0 cm long hish white abaxiali



Figure 1: Ficus deltoidea Jack plant.

Traditionally, *F. deltoidea* (Figure 1) has been used commonly as decorative houseplant and as traditional medicines [10] to heal ailments including sores, wounds, rheumatism, diabetes and as an after-birth tonic [11]. For many years, the dried leaves of *F. deltoidea* have been consumed and distributed among Malaysians as herbal tea and capsules. The decoction of the leaves is claimed to have antioxidant, antidiabetic, and aphrodisiac properties as well as in improving the blood circulation and in treating gout [11, 12, 13].



Figure 2: Andrographis paniculata plant.

A. paniculata (Figure 2) leaves have always been taken orally to reduce diabetes and high blood pressure. A. paniculata has a broad range of therapeutic properties and hence has been used for many years to treat upper gastrointestinal tract and upper respiratory infections, fever and Herpes [14]. Based on Roy et al., andrographolide is one of the bioactive components present in A. paniculata and has been reported for its anticancer [15], anti-HIV [16] and cardio protective properties among others [17].



Figure 3: Clinacanthus nutans plant.

In Malaysia, *C. nutans* (Figure 3) has been utilized traditionally to treat skin rashes, scorpion and insect bites; lesions caused by *Herpes simplex*, Diabetes mellitus, fever and are also used as diuretics [18, 19]. Some traditional practitioners in Malaysia boil the fresh leaves of *C. nutans* with water and consume as herbal tea [20]. *C. nutans* has been used as an antivenom, anti-inflammatory, analgesic, antidiabetic, anti-rheumatism, antiviral and as an antioxidant [21].



Figure 4: Cinnamomum verum plant.

Traditionally in Malaysia, stem bark of *C. verum* (Figure 4) is used to improve blood circulation, to make the body feel warm, to encourage contraction of the uterus, to expel wind, to prevent fits and as a laxative [22]. Besides that, the essential oils of *C. verum* are used in aromatherapy to ease emotional and mental fatigue [22]. The aromatic, astringent, expectorant and carminative properties of the bark of *C. verum* is used to treat stomach cramps, gastric irritation, dysentery, diarrhoea [23], neuralgia, rheumatism, toothache and paralysis of the tongue. *C. verum* stimulates the uterine muscles by reducing the

difficulties in deliveries due to inadequate contractions, promotes regular and easy menstruation and is taken as a warming herb for cold conditions. *C. verum* is also used as traditional remedy for aching muscles and other symptoms of viral conditions such as colds and flu [24].



Figure 5: Curcuma longa plant.

Traditionally, *C. longa* (Figure 5) has been applied in Malay traditional medicine for parturition, to treat amenorrhea, swelling, urogenital problems, wounds and as diuretic, lactagogue and tonic. Usually, hot water mixture of ground rhizome parts of *C. longa* is given orally to improve blood circulation, to regain body strength, to expel wind, to ease muscular and joint pain as well as to ease abdominal discomfort [22].

3 PHYTOCHEMICAL	COMPOUNDS	AND
PHYTOCHEMICAL	PROPERTIES	OF
MEDICINAL PLANTS		

3.1 Phytochemical compounds

Several studies on *F. deltoidea* leaves have been carried out to identify and isolate the phytochemical constituents of *F. deltoidea*. Study by Lip et al. using nuclear magnetic resonance (NMR) and mass spectrometers has identified moretenol ($C_{30}H_{50}O$) in *F. deltoidea* leaves [25].

Meanwhile, another study by Suryati et al. discovered an antibacterial compound known as lupeol ($C_{30}H_{50}O$) (Figure 6A) which exhibited toxicity against *Staphylococcus aureus*, *Bacillus subtilis*, and *Escherichia coli* [25, 26]. Also, the flavonoid compounds (rutin, quercetin and naringenin - Figure 6B, C, D) present in *F. deltoidea* leaves were confirmed by liquid chromatography-mass spectrometry (LC-MS) [27].



Figure 6: Phytochemical compounds present in *F. deltoidea* A: Lupeol; B: Rutin; C: Quercetin and D: Narigenin.



Figure 7: Phytochemical compounds present in *A. paniculata* A: Andrographolide; B: Deoxyandrographolide; C: Neoandrographolide and D: Dehydroandrographolide.

Based on previous studies, the aerial parts of *A. paniculata* (leaves and stems) were used in the extraction and isolation of the active phytochemical compounds [17, 28]. The major active phytochemical compound that was found in *A. paniculata* was andrographolide ($C_{20}H_{30}O_5$) (Figure 7A) [28].

Andrographolide is colourless, crystalline in appearance that has a bitter taste [15, 28] and exhibits therapeutic effects such as antiinflammatory, anti-microbial, anti-viral, immunostimulatory, anti-platelet aggregation [29, 30], anticancer, anti-HIV [14], cardio-protective and hepatoprotective effects [14, 17, 31]. Apart from that, other active phytochemical compounds present in A. paniculata are deoxyandrographolide $(C_{20}H_{30}O_4)$, neoandrographolide $(C_{26}H_{40}O_8)$ and dehydroandrographolide (C20H28O4) (Figure 7B, C, D-respectively) [32]. Dehydroandrographolide has properties such as hypotensive effect. vasorelaxant activity, anti-stimulant effect on production of nitric oxide and shows inhibition against human immuno-deficiency virus [30]. Also,

andrographolide, neoandrographolide, and dehydroandrographolide are reported to have virucidal properties against *Herpes simplex* virus type 1 (HSV-1) [33].

Teshima and colleagues isolated six Cglycosyl flavones from methanolic extract of *C. nutans* leaves including vitexin, isomollupentin 7-O- β -glucopyranoside, orientin, isoorientin, schaftoside (C₂₆H₂₈O₁₄) and isovitexin (Figure 8A, B, C, D, E, F-respectively) [34].



Figure 8: Six C-glycosyl flavones present in methanolic extract of C. nutans A: Vitexin; B: Isomollupentin 7-O- β -glucopyranoside; C: Orientin; D: Isoorientin; E: Schaftoside and F: Isovitexin.

These compounds possessed significant pharmacological properties such as antimicrobial (isoorientin and vitexin). hepatoprotective (isoorientin) and antioxidant activities (isovitexin) [34]. Gas chromatography mass spectrometry (GCMS) analysis of chloroform extract of C. nutans identified the major compound which was 1,2-benzenedicarboxylic acid, mono (2-ethylhexyl) ester. This phytochemical compound contributes to the medicinal activity and possesses antiantioxidant microbial, and antiproliferative properties [35]. Previous studies on the active phytochemical compounds of C. nutans extracts also discovered stigmasterol, β-sitosterol, lupeol [36] and betulin [37] (Figure 9A, B, C, D, Erespectively).

Previous study has shown that the major active phytochemical compounds in *C. verum* were eugenol ($C_{10}H_{12}O_2$) [38], linalool ($C_{10}H_{18}O$), cinnamaldehyde ($C_{9}H_{8}O$) [39], coumarin ($C_{9}H_{6}O_2$) [23] and benzyl benzoate ($C_{14}H_{12}O_2$) [40]. Coumarins are active compounds that can be found naturally in plants including *C. verum* and possess anticoagulant properties. High percentage of coumarin in plants can lead to health risks and become toxic to the liver if this compound is consumed in higher quantity on a regular basis.



Figure 9: Other phytochemical compounds present in *C. nutans* A: 1, 2-benzenedicarboxylic acid, mono (2- ethylhexyl) ester; B: stigmasterol; C: β -sitosterol and D: Lupeol and E: Betulin.

However, *C. verum* only contains low percentage of coumarin and still considered as safe and free from causing health risks [41]. The inhibitory activity of *C. verum* extract was attributed to the presence of cinnamaldehyde [42]. Previous studies have also shown that sensitive and resistant bacteria strain of *Helicobacter pylori* was completely inhibited by cinnamaldehyde compound [42, 43]. These active compounds of *C. verum* are very important as they possess important pharmacological properties as shown in Table III.



Figure 10: Phytochemical compounds present in C. longa A: α -turmerone; B: β -turmerone; C: aromatic-tumerone; D: aromatic-curcumene and E: alpha-santalene.

Based on previous study, the major phytochemical compounds that were identified in *C. longa* were α -turmerone (C₁₅H₂₂O) and β -turmerone [44] (Figure 10A, B-respectively). This study has also reported the presence of aromatic-turmerone, aromatic-curcumene and alpha-santalene (Figure 10 C, D, E-respectively) in dry rhizome oil [44]. The pharmacological properties of aromatic-turmerone are antimicrobial, larvicidal and antioxidant properties [45]. Turmerone possesses a variety of pharmacological activities such as antioxidant, anti-inflammatory, anti-tumor, anti-proliferative and anti-depressant activities [46].

Table III: Phytochemical compounds present in C. verum and their pharmacological properties



The major phenolic compounds identified in C. longa were curcuminids which can be divided into three types; curcumin (curcumin I). demethoxvcurcumin (curcumin II) and bisdemethoxycurcumin (curcumin III) [45]. These curcuminoids play an important role as anti-tumor agent [45] and are responsible for the yellow colour of turmeric [47]. Based on Nisar et al., [47] curcumin can be defined as yellow orange crystalline substance that is insoluble in water. Table IV shows curcuminoid derivatives and its pharmacological activities.



3.2 Phytochemical properties

3.2.1 F. deltoidea

F. deltoidea contains a lot of antioxidant compounds that possess various phytochemical properties such as antiangiogenic and anticancer properties. Angiogenesis is known as development of new blood vessels and has important roles in pathogenesis of various human diseases including cancer, psoriasis, arterial plague formation, ocular neovascularization, gastrointestinal ulcers, rheumatoid arthritis, and diabetic retinopathy. Shafaei et al. reported that the antiangiogenic effect of F. deltoidea extract was due to the presence of relatively high contents of ursolic acid, phenolics and flavonoids. The selective cytotoxicity towards colon and breast cancer cell lines, and anti-angiogenic effect indicated the potential anti-cancer effect of F. deltoidea extracts [13]. Investigation on the cytotoxicity of aqueous and ethanolic plant extracts of *F. deltoidea* on human carcinoma cells revealed that both extracts could cause apoptosis at a concentration of 1000 µg/ml. An aqueous extract of F. deltoidea promoted cell detachment while the ethanolic tried to inhibit cell proliferation through DNA fragmentation [48].

Apart from that, the methanol extract of F. deltoidea showed potential as an antidiabetic agent by inhibiting the hepatic glucose production and promoting glucose utilization [49]. Farsi et al. evaluated the enzymes inhibitory effect and antioxidant properties of different fractions of methanolic extract obtained from F. deltoidea leaves. The n-butanol fraction revealed significant α -glucosidases and α -amylase inhibitory effects (IC50 values of 15.1 and 39.42 ug/ml. respectively) along with the remarkable antioxidant activity when compared to the other fractions and indicated that F. deltoidea could be a potential source of promising anti-diabetic drug [49]. Adam and colleagues evaluated the potential of five extracts and three fractions of *F. deltoidea* to enhance basal and insulin-stimulated glucose uptake into the Chang liver cell line and found that all F. deltoidea extracts and fractions except for ether extract possessed the ability to enhance either the basal or insulin-stimulated glucose uptake into this cell line [50]. One of the therapeutic techniques to control postprandial hyperglycaemia is by the inhibition of carbohydrate hydrolysing enzymes such as α-glucosidases and α -amylase [49]. Another study assessed on the ability of the crude extracts and fractions of F. deltoidea as antidiabetic agent to inhibit yeast and

mammalian α -glucosidase as well as α -amylase. The results suggested that the crude extracts and fractions of *F. deltoidea* inhibited both yeast and rat intestinal α -glucosidases in a dose-dependent manner. However, the extracts and fractions of *F. deltoidea* did not inhibit porcine pancreatic α -amylase [51].

Study by Abdullah et al. suggested standardization of extracts of different varieties of F. deltoidea for anti-inflammatory activity using three in vitro assays: lipoxygenase, hyaluronidase, and TPA-induced oedema. The results of the different extracts from three varieties of the plant showed anti-inflammatory activities [52]. Evaluation of the antinociceptive activity of F. deltoidea aqueous extract by using three models of nociception; acetic acid-induced abdominal writhing, formalin and hot plate test revealed that F. deltoidea leaves' aqueous extract contained pharmacologically active constituents that possessed antinociceptive activity justifying its traditional therapeutic use in treating conditions related with painful conditions [53]. Study on the anti-inflammatory activity of the aqueous extract of F. deltoidea in rats using a carrageenan-induced paw oedema test, a cotton pellet-induced granuloma test, and a formalin test showed that there was significant anti-inflammatory activity in every test with a dose-response effect [54].

3.2.2 A. Paniculata

Study by Al-Henhena and colleagues revealed that A. paniculata possesses antioxidant, antiproliferative and antimetastatic properties and act as free radical scavengers as the ethanol extract of A. paniculata interferes with the intermediate biomarker for colon cancer development. They also suggested that A. paniculata has the potential to be a new anti-cancer therapeutic agent due to its diterpenoid and flavonoid contents. Previously, diterpenoids showed potent effect on the inhibition of HT29 (colon cancer cell line) proliferation [55]. Previous researchers also have discovered the antioxidant and gastro protective activities of aqueous and ethanol extracts of A. paniculata in Sprague Dawley rats by pre-treating the rats with the extracts where they used carboxymethyl cellulose as negative control and omeprazole as the positive control. The results showed that there was a significant dose dependent reduction in gastric lesions with increased pH and mucus content of stomach in the rats pre-treated with omeprazole and both the extracts [56]. They concluded that antioxidant is the main factor that protects the cells from damage caused by oxidative stress [56]. A study on *A. paniculata* aerial parts in 20 patients with type 2 diabetes mellitus for 12 weeks exhibited antidiabetic effects with no significant adverse effects on the patients [57]. Previous study reported that andrographolide and ethanol extract of *A. paniculata* possessed antidiabetic, hypolidemic and antioxidant properties in adult streptozotocin-nicotinamide type 2 diabetes mellitus (STZ-NA T2DM) rats [58].

Malahubban and team discovered the antibacterial activities of ethanol, methanol and aqueous extracts of A. paniculata against enterica, Esherichia Salmonella coli, Staphylococcus aureus and Bacillus cereus. Methanol extract was found to be the most effective against all the bacteria compared to other extracts due to its polarity [59]. Uridine diphosphate glucuronosyltransferase (UGT) is an enzyme required in glucuronidation process. Ahmad and colleagues studied the effects of A. paniculata extract on UGT activity and suggested that the ethanol extract of A. paniculata at 500 µg/mg protein significantly reduced the UGT activity [60]. Recent study on the effect of A. paniculata extract in Malaysia on the activity of cDNA-expressed UGT isoforms was conducted where the results showed that it inhibited UGT1A and UGT2B isoenzymes. This suggests that A. paniculata has the potential for drug-herbal extract interactions in the therapeutic setting [61]. Ethanol extract of A. paniculata was reported to influence the restoration of different enzyme levels of serum glutamine pyruvate transaminase (SGPT), serum glutamine oxaloacetate transaminase (SGOT), gamma-glutamyl transpeptidase (GGTP) and serum alkaline phosphatase (SALP) after carbon tetrachloride- (CCL4-) induced liver injury [62, 63]. Study on the effect of andrographolide and the leaf extract of A. paniculata against tetrachloride (CCL4-) induced hepatic microsomal lipid peroxidation in vitro was completely protected by the leaf extract but not by andrographolide, indicating that the hepatoprotective effect is not solely due to the presence of andrographolide [63, 64]. Other compounds of A. paniculata extracts neoandrographolide such as and dehydroandrographolide have also been reported to possess hepatoprotective effect [63].

3.2.3 Clinacanthus nutans

C. nutans plants possess variety of pharmacological properties including anti-venom, anti-inflammatory, analgesic, antidiabetic, anti-

rheumatism, antiviral and antioxidant properties. Investigation of *C. nutans* (chloroform, methanol and water) extracts on diphenyl-1-picrylhydrazyl (DPPH), galvinoxyl radical, nitric oxide, and hydrogen peroxide scavenging assays revealed that *C. nutans* extracts contained antioxidant agents that were capable of negating free radicals [35]. However, the antioxidant activities of the extract varied for each test due to the solubility of the extracts in different testing systems and the stereo selectivity of the radicals [35].

Saad and colleagues investigated the ethanolic extract of C. nutans on the standard free radical, 2,2-diphenyl-1-picyrl-hydrazyl (DPPH) and the result showed that C. nutans possessed antioxidant activity [65]. Recent study suggested that C. nutans has anticancer properties as the IC50 value was potent enough to inhibit the growth of cancer cells [66]. Moreover, C. nutans contains low levels of polyamine and has anti-proliferative effect on human lung adenocarcinoma cell line (A549 cell) [66]. Another study by Murni and team led to the finding of anti-proliferative effect of C. nutans on human ovarian cancer cell line (SKOV-3), breast cancer cell line (MCF-7) and human colorectal adenocarcinoma (HT-29) [67]. Yahaya et al. (2015) claimed C. nutans as a plant with high medicinal values that possessed potential anticancer. antioxidant. antidiabetic. immunomodulatory, wound healing, antiinflammatory and analgesic activities [68].

3.2.4 Cinnamomum verum

For many years, *C. verum* has been used widely all around the world in culinary as spices as well as in therapeutic treatments. Previous studies on the bark and leaves of *C. verum* have reported various important pharmacological properties such as antidiabetic, antinociceptive, astringent, diuretic activities and antifungal activities (69). An earlier study suggested that a compound cinnamtannin B1, isolated from the stem bark of *C. verum* might have insulin-like activity. Cinnamtannin B1 can be used as a drug to initiate insulin receptor signalling by stimulation of phosphorylation that will be useful in the treatment of type 2 diabetes [70].

Another study by Azahari and team suggested that the extract of cinnamon showed insulin-mimicking effects in adipocytes. They revealed that cinnamon enhances the glucose uptake, reduces the lipid breakdown and resembles insulin activity [71]. The oils, leaves and barks of *C. verum* also showed highest

antimicrobial activities against Candida albicans, Candida parapsilosis, Candida tropicalis, Candida glabrata, Cryptococcus neoformans, Aspergillus fumigatus, Trichophyton rubrum, Trichophyton mentagrophytes, Trichophyton tonsurans. Microsporum gypseum, Microsporum audouinii and Microsporum canis. Bark and leaf oils of C. verum showed strong antimicrobial activity due to the presence of cinnamaldehyde and eugenol [72]. C. verum exhibited activities against neurological disorders such as Parkinson's and Alzheimer's and possessed antioxidant, anti-inflammatory, antidiabetic. antimicrobial. anti-cancer. lipidlowering and cardiovascular-disease-lowering compounds [73].

3.2.5 Curcuma longa

The phytochemical compounds present in C. longa possess cardio-protective, antianti-tumor, inflammatory. antifungal, immunomodulatory, antioxidation, antimutagenic activities, has a protective effect against aflatoxin B1 (AFB1) induced toxicity, antibacterial activities and anti-human immunodeficiency virus activity [74]. Previous study showed that curcumin from C. longa has the capability to delay the inflammatory response and reduced the occurrence of the joint inflammation symptoms [75]. Zahidah and team concluded that oral consumption of curcumin at the dose of 100 mg/ml/kg/day for 28 days (collagen-induced arthritis CurcuminT) and 42 days (collagen induced arthritis CurcuminC) have a potential in delaying and improving joint abnormality and injury in Sprague-Dawley rats with collagen induced arthritis.

Hussan and his team suggested curcumin as an alternative agent for oestrogen for prevention of postmenopausal osteoporosis [76]. A study regarding curcumin from C. longa showed that that curcumin exhibited cytotoxic effect towards acute myeloblasticleukaemia (HL-60), breast adenocarcinoma (MCF-7), chronic myelogenicleukaemia (K-562) and cervical epithelial carcinoma (HeLa). This study suggested that curcumin can act as a potential cancer controlling agent by the induction of apoptosis in breast cancer [77]. Another study showed that ethanolic extract of C. longa exhibited anthelmintic properties at 200 mg/ml against Haemonchus larvae and can be used as a substitute for levamisole [78]. Khaliq et al. in their review suggested that the extracts of C. longa showed hypoglycaemic and hypolipidaemic effects and that further studies were needed to investigate the mechanisms for their hypoglycaemic potential. *C. longa* also reduced the blood glucose level and had useful effects against diabetic complications [79].

4 CONCLUSIONS

Medicinal plants are still being utilised by traditional practitioners as folk medicines as remedies in treating various diseases rather than using the conventional medicines. This is due to the presence of beneficial chemical compounds that have healing properties in addition to combating microbial infections and other diseases. This review article highlights on the phytochemical properties present in five medicinal plants in Malaysia. Most of the medicinal plants have activities such as antimicrobial, anticancer, antidiabetic and anti-inflammatory properties though some have their own traditional functions as an afterbirth tonic and as an additive in energy drinks. The current research trends in Malaysia involve exploring the herbal plants in terms of anticancer. antidiabetic, anti-inflammatory, antimicrobial, anti-proliferative, antioxidant and antinociceptive properties. Hence, the phytochemical properties that are exhibited by these medicinal plants should be focused in finding new drugs in economical ways.

CONFLICT OF INTEREST

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

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REFERENCES

- Parivuguna V. Antimicrobial properties and phytochemical constituents of Rhoeo discolor Hance. Ethnobotanical Leaflets 2008; 12: pp 841-845.
- [2] Njoku OV, Obi C. Phytochemical constituents of some selected medicinal plants. African Journal of Pure and Applied Chemistry 2009; 3(11): pp 228-233.
- [3] Wadood A, Ghufran M, Jamal S, Naeem M, Khan A, Ghaffar R. Phytochemical analysis of medicinal plants occurring in local area of Mardan. Biochemistry & Analytical Biochemistry 2013; 2(144): pp 2-4.

- [4] J. Harborne, "Phytochemical Methods," in A guide to modern techniques of plant analysis, 3th ed. London, UK: Chapman & Hall, 1973, pp 52-114.
- [5] Aberoumand A. Screening of phytochemical compounds and toxic proteinaceous protease inhibitor in some lesserknown food based plants and their effects and potential applications in food. International Journal of Food Science and Nutrition Engineering 2012; 2(3): pp 16-20.
- [6] Kennedy DO, Wightman EL. Herbal extracts and phytochemicals: plant secondary metabolites and the enhancement of human brain function. Advances in Nutrition: An International Review Journal 2011; 2(1): pp 32-50.
- [7] Pandey KB, Rizvi SI. Plant polyphenols as dietary antioxidants in human health and disease. Oxidative Medicine and Cellular Longevity 2009; 2(5): pp 270-278.
- [8] Hussain MS, Fareed S, Saba Ansari M, Rahman A, Ahmad IZ, Saeed M. Current approaches toward production of secondary plant metabolites. Journal of Pharmacy & Bioallied Sciences 2012; 4(1): pp 10-20.
- [9] Savithramma N, Rao ML, Suhrulatha D. Screening of medicinal plants for secondary metabolites. Middle-East Journal of Scientific Research 2011; 8(3): pp 579-584.
- [10] Fatihah HNN, Mat N, Zaimah AR, Zuhailah MN, Norhaslinda H, Khairil M, et al. Morphological phylogenetic analysis of seven varieties of *Ficus deltoidea* Jack from the Malay Peninsula of Malaysia. PLoS ONE 2012; 8(8): pp 10.
- [11] Bunawan H, Amin N, Bunawan S, Baharum S, Noor N. Ficus deltoidea Jack: A review on its phytochemical and pharmacological importance. Evidence-Based Complementary and Alternative Medicine 2014; vol. 2014: pp 1-8.
- [12] Fatihah HNN, Nashriyah M, Zaimah ARN, Khairil M, Ali AM. Leaf morphology and anatomy of 7 varieties of *Ficus deltoidea* (Moraceae). Turkish Journal of Botany 2014; 38(4): pp 677-685.
- [13] Shafaei A, Muslim NS, Nassar ZD, Aisha AF, Majid AMSA, Ismail Z. Antiangiogenic effect of *Ficus deltoidea* Jack standardised leaf extracts. Tropical Journal of Pharmaceutical Research 2014; 13(5): pp 761-768.
- [14] Roy S, Rao K, Bhuvaneswari C, Giri A, Mangamoori LN. Phytochemical analysis of *Andrographis paniculata* extract and its antimicrobial activity. World Journal of Microbiology and Biotechnology 2010; 26(1): pp 85.
- [15] Sheeja K, Kuttan G. Activation of cytotoxic T lymphocyte responses and attenuation of tumor growth in vivo by *Andrographis paniculata* extract and andrographolide. Immunopharmacology and Immunotoxicology 2007; 29(1): pp 81-93.
- [16] Calabrese C, Berman SH, Babish JG, Ma X, Shinto L, Dorr M, et al. A phase I trial of andrographolide in HIV positive patients and normal volunteers. Phytotherapy Research 2000; 14(5): pp 333-338.
- [17] Yoopan N, Thisoda P, Rangkadilok N, Sahasitiwat S, Pholphana N, Ruchirawat S, et al. Cardiovascular effects of 14-deoxy-11, 12-didehydroandrographolide and Andrographis paniculata extracts. Planta Medica 2007; 73(06): pp 503-511.
- [18] Lau K, Lee S, Chin J. Effect of the methanol leaves extract of *Clinacanthus nutans* on the activity of acetylcholinesterase in male mice. Journal of Acute Disease 2014; 3(1): pp 22-25.
- [19] Ghasemzadeh A, Nasiri A, Jaafar HZ, Baghdadi A, Ahmad I. Changes in phytochemical synthesis, chalcone synthase activity and pharmaceutical qualities of Sabah snake grass (*Clinacanthus nutans* L.) in relation to plant age. Molecules 2014; 19(11): pp 17632-17648.

[22]

- [20] Aslam MS, Ahmad MS, Mamat AS. A Review on Phytochemical Constituents and Pharmacological Activities of *Clinacanthus nutans*. International Journal of Pharmacy and Pharmaceutical Sciences 2014; 7(2): pp 30-33.
- [21] Arullappan S, Rajamanickam P, Thevar N, Kodimani CC. In vitro screening of cytotoxic, antimicrobial and antioxidant activities of *Clinacanthus nutans* (Acanthaceae) leaf extracts. Tropical Journal of Pharmaceutical Research 2014; 13(9): pp 1455-1461.
- [22] Jamal JA, Ghafar ZA, Husain K. Medicinal plants used for postnatal care in Malay traditional medicine in the Peninsular Malaysia. Pharmacognosy Journal 2011; 3(24): pp 15-24.
- [23] Hema R, Kumaravel S, Devi Martina T. Chromatograph interfaced to a mass spectrometer analysis of *Cinnamomum verum*. Nature and Science 2010; 8(11): pp 152-155.
- [24] Husain SS, Ali M. Analysis of volatile oil of the stem bark of *Cinnamomum zeylanicum* and its antimicrobial activity. International Journal of Research in Pharmacy & Science 2013; 3(4): pp 40-49.
- [25] Lip JM, Hisham DN, Zaidi JA, Musa Y, Ahmad A, Normah A, et al. Isolation and identification of moretenol from *Ficus deltoidea* leaves. Journal of Tropical Agriculture and Food Science 2009; 37(2): pp 195-201.
- [26] Suryati S, Nurdin H, Dachriyanus D, Lajis MNH. Structure elucidation of antibacterial compound from *Ficus deltoidea* Jack leaves. Indonesian Journal of Chemistry 2011; 11(1): pp 67-70.
- [27] Ong S, Ling A, Poospooragi R, Moosa S. Production of flavonoid compounds in cell cultures of *Ficus deltoidea* as influenced by medium composition. International Journal of Medicinal Aromatic Plants 2011; 1(2): pp 62-74.
- [28] Yadav J, Singh T. Phytochemical analysis and antifungal activity of *Andrographis paniculata*. International Journal of Pharmaceutical Research and Bio Science 2012; 1(4): pp 240-263.
- [29] Kumar RA, Sridevi K, Kumar NV, Nanduri S, Rajagopal S. Anticancer and immunostimulatory compounds from Andrographis paniculata. Journal of Ethnopharmacology 2004; 92(2): pp 291-295.
- [30] Xu T, Pan J, Zhao L. Simultaneous determination of four andrographolides in *Andrographis paniculata* Nees by silver ion reversed-phase high-performance liquid chromatography. Journal of Chromatographic Science 2008; 46(8): pp 747-750.
- [31] Jayakumar T, Hsieh CY, Lee JJ, Sheu JR. Experimental and clinical pharmacology of *Andrographis paniculata* and its major bioactive phytoconstituent andrographolide. Evidence-Based Complementary and Alternative Medicine 2013; vol. 2013: pp 16.
- [32] Chao W-W, Lin B-F. Review isolation and identification of bioactive compounds in *Andrographis paniculata* (Chuanxinlian). Chinese Medicine 2010; 10: pp 44.
- [33] Akbar S. Andrographis paniculata: A review of pharmacological activities and clinical effects. Alternative Medicine Review 2011; 16(1): pp 66-77.
- [34] Teshima K, Kaneko T, Ohtani K, Kasai R, Lhieochaiphant S, Picheansoonthon C, et al. C-Glycosyl Flavones from *Clinacanthus nutans* (Natural Medicine Note). Natural Medicines 1997; 51(6): pp 557.
- [35] Yong YK, Tan JJ, Teh SS, Mah SH, Ee GCL, Chiong HS, et al. *Clinacanthus nutans* extracts are antioxidant with antiproliferative effect on cultured human cancer cell lines. Evidence-Based Complementary and Alternative Medicine 2013; vol. 2013: pp 8.

- [36] Dampawan P, Huntrakul C, Reutrakul V, Raston CL, White AH. Constituents of *Clinacanthus nutans* and the crystal structure of lup-20(29)-ene-3-one. Journal of the Science Society of Thailand 1977; 1(2): pp 53-62.
- [37] Lin J, Li H, Yu J. Studies on the chemical constituents of niu xu hua (*Clinacanthus nutans*). Zhongcaoyao 1983; 14: pp 337-338.
- [38] Senanayake UM, Lee TH, Wills RB. Volatile constituents of cinnamon (*Cinnamomum zeylanicum*) oils. Journal of Agricultural and Food Chemistry 1978; 26(4): pp 822-824.
- [39] Variyar P, Bandyopadhyay C. On Some Chemical Aspects of *Cinnamomum zeylanicum*. Pafai Journal 1989; 10(4): pp 35-38.
- [40] Rao Y, Paul S, Dutta P. Major constituents of essential oils of *Cinnamomum zeylanicum*. Indian Perfumer 1988; 32(1): pp 86-89.
- [41] Ranasinghe P, Pigera S, Premakumara GS, Galappaththy P, Constantine GR, Katulanda P. Medicinal properties of 'true' cinnamon (*Cinnamomum zeylanicum*): a systematic review. BMC Complementary and Alternative Medicine 2013; 13(1): pp 275.
- [42] Uma B, Prabhakar K, Rajendran S, Lakshmi Sarayu Y. Studies on GC/MS spectroscopic analysis of some bioactive antimicrobial compounds from *Cinnamonum zeylanicum*. Journal of Medicinal Plants 2009; 3(31): pp 125-131.
- [43] Nir Y, Potasman I, Stermer E, Tabak M, Neeman I. Controlled trial of the effect of cinnamon extract on *Helicobacter pylori*. Helicobacter 2000; 5(2): pp 94-97.
- [44] Singh G, Kapoor I, Singh P, De Heluani CS, De Lampasona MP, Catalan CA. comparative study of chemical composition and antioxidant activity of fresh and dry rhizomes of turmeric (*Curcuma longa* Linn.). Food and Chemical Toxicology 2010; 48(4): pp 1026-1031.
- [45] Li S, Yuan W, Deng G, Wang P, Yang P, Aggarwal B. Chemical composition and product quality control of turmeric (*Curcuma longa* L.). Pharmaceutical Crops 2011; 2: pp 28-54.
- [46] Liao JC, Tsai JC, Liu CY, Huang HC, Wu LY, Peng WH. Antidepressant-like activity of turmerone in behavioral despair tests in mice. BMC Complementary and Alternative Medicine 2013; 13(1): pp 299.
- [47] Nisar T, Iqbal M, Raza A, Safdar M, Iftikhar F, Waheed M. Turmeric: A promising spice for phytochemical and antimicrobial activities. American-Eurasian Journal of Agricultural & Environmental Sciences 2015; 15(7): pp 1278-1288.
- [48] Akhir NAM, Chua LS, Majid FAA, Sarmidi MR. Cytotoxicity of aqueous and ethanolic extracts of *Ficus deltoidea* on human ovarian carcinoma cell line. British Journal of Medicine and Medical Research 2011; 1(4): pp 397.
- [49] Farsi E, Shafaei A, Hor SY, Ahamed MBK, Yam MF, Attitalla IH, et al. Correlation between enzymes inhibitory effects and antioxidant activities of standardized fractions of methanolic extract obtained from *Ficus deltoidea* leaves. African Journal of Biotechnology 2011; 10(67): pp 15184-15194.
- [50] Adam Z, Hamid M, Ismail A, Khamis S. Effect of *Ficus deltoidea* extracts on hepatic basal and insulin-stimulated glucose uptake. Journal of Biological Sciences 2009; 9(8): pp 796-803.
- [51] Misbah H, Aziz AA, Aminudin N. Antidiabetic and antioxidant properties of *Ficus deltoidea* fruit extracts and fractions. BMC Complementary and Alternative Medicine 2013; 13(1): pp 118.
- [52] Abdullah Z, Hussain K, Ismail Z, Ali RM. Anti-inflammatory activity of standardised extracts of leaves of three varieties

of *Ficus deltoidea*. Asian Journal of Pharmaceutical and Clinical Research 2009; 1(3): pp 100-105.

- [53] Sulaiman MR, Hussain M, Zakaria ZA, Somchit M, Moin S, Mohamad A, et al. Evaluation of the antinociceptive activity of *Ficus deltoidea* aqueous extract. Fitoterapia 2008; 79(7): pp 557-561.
- [54] Zakaria Z, Hussain M, Mohamad A, Abdullah F, Sulaiman M. Anti-inflammatory activity of aqueous extract of *Ficus deltoidea*. Biological Research for Nursing 2011; 14(1): pp 90-97.
- [55] Al-Henhena N, Ying RPY, Ismail S, Najm W, Khalifa SA, El-Seedi H, et al. Chemopreventive efficacy of *Andrographis paniculata* on azoxymethane-induced aberrant colon crypt foci in vivo. PloS one 2014; 9(11): pp 1-12.
- [56] Wasman S, Mahmood A, Chua LS, Alshawsh MA, Hamdan S. Antioxidant and gastroprotective activities of Andrographis paniculata (Hempedu Bumi) in Sprague Dawley rats. Indian Journal of Experimental Biology 2011; vol.49: pp 767-772.
- [57] Agarwal R, Sulaiman SA, Mohamed M. Open label clinical trial to study adverse effects and tolerance to dry powder of the aerial part of Andrographis *paniculata* in patients type 2 with *Diabetes mellitus*. Malaysian Journal of Medical Sciences 2005; 12(1): pp 13-19.
- [58] Subramanian R, Asmawi M, Sadikun A. Effect of andrographolide and ethanol extract of *Andrographis paniculata* on liver glycolytic, gluconeogenic, and lipogenic enzymes in a type 2 diabetic rat model. Pharmaceutical Biology 2008; 46(10-11): pp 772-780.
- [59] Malahubban M, Alimon AR, Sazili AQ, Fakurazi S, Zakry FA. Phytochemical analysis of Andrographolis paniculata and Orthosipon stamineus leaf extracts for their antibacterial and antioxidant potential. Tropical Biomedicine 2013; 30(3): pp 467-480.
- [60] Ahmad N, Ismail S, Hussin AH, Syahrani A, Palupi T, Triyanti UF, et al. Effects of *Andrographis paniculata* extract on uridine diphosphate-glucuronosyl transferase activity. Malaysian Journal of Pharmaceutical Sciences 2005; 3(1): pp 25-33.
- [61] Ismail S, Aziah Hanapi N, Ab Halim MR, Uchaipichat V, Mackenzie PI. Effects of Andrographis paniculata and Orthosiphon stamineus extracts on the glucuronidation of 4-methylumbelliferone in human UGT isoforms. Molecules 2010; 15(5): pp 3578-3592.
- [62] Verma VK, Sarwa KK, Kumar A, Zaman MK. Comparison of hepatoprotective activity of Swertia chirayita and Andrographis paniculata plant of North–East India against CCI 4 induced hepatotoxic rats. Journal of Pharmacy Research 2013; 7(7): pp 647-653.
- [63] Hossain MS, Urbi Z, Sule A, Rahman K. Andrographis paniculata (Burm. f.) Wall. ex Nees: A review of ethnobotany, phytochemistry, and pharmacology. The Scientific World Journal 2014; vol.2014: pp 28.
- [64] Choudhury B, Poddar M. Andrographolide and kalmegh (Andrographis paniculata) extract: in vivo and in vitro effect on hepatic lipid peroxidation. Methods and Findings in Experimental and Clinical Pharmacology 1984; 6(9): pp 481-485.
- [65] Saad R, Pohyeen T, Khan J, Wenji L, Sultan S, Abdul Hameed J, et al. Phytochemical screening and antioxidant activity of different parts from five Malaysian herbs. International Journal of Science and Technology 2014; 19(2): pp 1336-1347.
- [66] Abdul Ghani R, Jamil EF, Nor Azahan Shah NAM, Nik Abdul Malek NN. The role of polyamines in antiproliferative effect of selected Malaysian herbs in human

lung adenocarcinoma cell line. Jurnal Teknologi (Sciences & Engineering) 2015; 77(25): pp 136-140.

- [67] Murni YN, Ling SK, Syarifah SS, Zunoliza A, Afiedatul MN. (2017, November). Anti-proliferative effect of *Clinacanthus nutans* on ovarian, breast and colorectal cancer cell lines. Presented at Persidangan Industri Herba.
- [68] Yahaya R, Dash GK, Abdullah MS, Mathews A. *Clinacanthus nutans* (burm. F.) Lindau: An useful medicinal plant of South-East Asia. International Journal of Pharmacognosy and Phytochemical Research 2015; 7(6): pp 1244-1250.
- [69] Joshi K, Awte S, Bhatnagar P, Walunj S, Gupta R, Joshi S, et al. *Cinnamomum zeylanicum* extract inhibits proinflammatory cytokine TNF: *in vitro* and *in vivo* studies. Research in Pharmaceutical Biotechnology 2010; 2(2): pp 14-21.
- [70] Taher M, Majid FAA, Sarmidi MR. A proanthocyanidin from *Cinnamomum zeylanicum* stimulates phosphorylation of insulin receptor in 3t3-L1 adipocytes. Jurnal Teknologi 2006; 44: pp 53-68.
- [71] Azahari N, Khattak MMAK, Taher M, Arief Ichwan S. Dose water extract of Cinnamon (*Cinnamomum zeylanicum*) exhibits anti-diabetic properties in cultured 3T3-L1 adipocytes: a concurrent assessment of adipogenesis, lipolysis and glucose uptakes. Journal of Food and Nutrition Research 2014; 2(11): pp 764-769.
- [72] Jantan IB, Karim Moharam BA, Santhanam J, Jamal JA. Correlation between chemical composition and antifungal activity of the essential oils of eight Cinnamomum species. Pharmaceutical Biology 2008; 46(6): pp 406-412.
- [73] Rao PV, Gan SH. Cinnamon: A multifaceted medicinal plant. Evidence-Based Complementary and Alternative Medicine 2014; vol.2014: pp 12.
- [74] Rajesh H, Rao S, Megha Rani N, Prathima K, Rajeesh E, Chandrashekar R. Phytochemical analysis of methanolic extract of *Curcuma longa* Linn Rhizome. International Journal of Universal Pharmacy and Bio Sciences 2013; 2(2): pp 39-45.
- [75] Zahidah A, Faizah O, Aqilah KN, Anna KT. Curcumin as an anti-arthritic agent in collagen-induced arthritic Sprague-Dawley rats. Sains Malaysiana 2012; 41(5): pp 591-595.
- [76] Hussan F, Ibraheem NG, Kamarudin TA, Shuid AN, Soelaiman IN, Othman F. Curcumin protects against ovariectomy-induced bone changes in rat model. Evidence-Based Complementary and Alternative Medicine 2012; vol.2012: pp 1-7.
- [77] Latifah S, Faujan H, Sze L, Raha A, Hisyam A, LI O. Curcumin from turmeric (*Curcuma longa*) induced apoptosis in human mammary carcinoma cells (MDA-MB-231). Malaysian Journal of Medicine and Health Sciences 2006; 2(2): pp 71-79.
- [78] Nasai NB, Abba Y, Abdullah FFJ, Marimuthu M, Tijjani A, Sadiq MA, et al. *In vitro* larvicidal effects of ethanolic extract of *Curcuma Longa* Linn. on *Haemonchus larval* stage. Veterinary World 2016; 9(4): pp 417-420.
- [79] Khaliq T, Sarfraz M, Ashraf M. Recent progress for the utilization of *Curcuma longa*, *Piper nigrum* and *Phoenix dactylifera* seeds against type 2 diabetes. The West Indian Medical Journal 2015; 64(5): pp 527-532.
- [80] Jarukamjorn K, Nemoto N. Pharmacological aspects of Andrographis paniculata on health and its major diterpenoid constituent andrographolide. Journal of Health Science 2008; 54(4): pp 370-381.
- [81] Kunsorn P, Ruangrungsi N, Lipipun V, Khanboon A, Rungsihirunrat K. The identities and anti-herpes simplex virus activity of *Clinacanthus nutans* and *Clinacanthus*

siamensis. Asian Pacific Journal of Tropical Biomedicine 2013; 3(4): pp 284-290.

- [82] Chakraborty A, Sankaran V, Ramar M, Chellappan DR. Chemical analysis of leaf essential oil of *Cinnamomum verum* from Palni Hills, Tamil Nadu. Journal of Chemical and Pharmaceucal Sciences 2015; 8(3): pp 476-479.
 [83] Kuddus MR, Rumi F, Kaisar MA, Hasan CM.
- [83] Kuddus MR, Rumi F, Kaisar MA, Hasan CM. Sesquiterpene and phenylpropanoids from *Curcuma longa*. Bangladesh Pharmaceutical Journal 2010; 13(2): pp 31-34.